Omega-3 Fatty Acids as a Putative Treatment for Traumatic Brain Injury

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Abstract

Traumatic brain injury (TBI) is a global public health epidemic. In the US alone, more than 3 million people sustain a TBI annually. It is one of the most disabling injuries as it may cause motor and sensory deficits and lead to severe cognitive, emotional, and psychosocial impairment, crippling vital areas of higher functioning. Fueled by the recognition of TBI as the "signature injury" in our wounded soldiers in Iraq and Afghanistan, and its often devastating impact on athletes playing contact sports, interest in TBI and TBI research has increased dramatically. Unfortunately, despite increased awareness of its detrimental consequences, there has been little progress in developing effective TBI interventions. Recent evidence, however, strongly indicates that nutritional intervention may provide a unique opportunity to enhance the neuronal repair process after TBI. To date, two omega-3 fatty acids, docosahexaenoic acid (DHA) and eicosapentaenoic acid (EPA), have the most promising laboratory evidence for their neuro-restorative capacities in TBI. Although both animal models and human studies of brain injuries suggest they may provide benefits, there has been no clinical trial evaluating the effects of n-3 fatty acids on resilience to, or treatment, of TBI. This article reviews the known functions of n-3 fatty acids in the brain and their specific role in the cellular and biochemical pathways underlying neurotraumatic injury. We also highlight recent studies on the therapeutic impact of enhanced omega 3 intake *in vivo*, and how this may be a particularly promising approach to improving functional outcome in patients with TBI.

Key words: encephalopathy; omega 3 fatty acids, plasma membrane; therapeutic approaches to CNS injury; traumatic brain injury

Introduction

RAUMATIC BRAIN INJURY (TBI) remains a significant cause of death and permanent disability in the United States. Drawing data from hospitalizations and emergency department visits only, the Centers for Disease Control and Prevention (CDC) estimates that 1.7 million people in the United States sustain a TBI each year. Approximately 15%–20% of U.S. soldiers in Iraq and Afghanistan also experience a TBI while deployed, making TBI one of the most common injuries among military personnel. It is estimated that 1.6–3.8 million sports-related TBIs occur in the United States annually, including those not treated by a health care provider. Seventy-five to eighty percent of TBIs, however, are mild, involving only a brief alteration in consciousness or mental status. Emerging research on the long-term effects of mild traumatic brain injury (mTBI) has drawn intense media attention and even Congressional scrutiny.

Repetitive head impacts add another level of complexity to the characterization of TBI because the emergence and duration of pathogenic events can overlap. This is particularly relevant in athletes and military personnel. Recent evidence suggests that chronic repetitive subconcussive head impacts may also result in cumulative long-term deleterious effects. Therefore, the summed effects of both concussive and subconcussive injuries may better represent the more complicated clinical landscape for TBI

TBI represents both an acute and chronic process. Although the immediate consequences of brain injury can be devastating, long-term health disorders associated with TBI include post-traumatic stress disorder (PTSD), neurodegenerative diseases (Alzheimer's disease or Parkinsonism), neurocognitive deficits, psychosocial health problems (e.g., binge drinking, major depression, impairment of social functioning and ability to work, suicide), epilepsy, pain, and other alterations in personality or behavior. 6–8

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TBI is a multifaceted disease with prolonged secondary pathogenesis and long-lasting adverse neurological sequelae that remain a clinical challenge to manage. Interventions targeting the acute phase of TBI, such as prevention of hypoxia and excitotoxicity, will differ from those targeting the chronic phase of TBI. Phase III clinical trials thus far have failed to yield an effective pharmacological strategy for neuroprotection in TBI, and this may be partly due to the use of drugs that target only a single pathophysiological pathway rather than the multiple mechanisms involved in secondary injury post-TBI. Targeting the multiple pathways that contribute to a deleterious secondary cascade may result in more successful clinical outcomes.

A growing body of preclinical data has shown that nutritional intervention, such as dietary supplementation with n-3 (also known as omega-3) fatty acids, may be of therapeutic benefit in acute injury to the brain. ^{10–13} Omega-3 fatty acids have long been known to play a restorative role in several pathways implicated in traumatic insult to the brain. 12,14–17 Emerging clinical evidence from both animal models and human studies of other brain injuries continue to suggest that they may provide benefits; however, there has been no human trial evaluating the effects of n-3 fatty acids on resilience to or treatment of TBI, ¹⁷–19 though there have been case studies on the use of omega-3s in the acute phase of severe head injury. 188,189 This article reviews the physiological functions of n-3 polyunsaturated fatty acids (PUFAs) in the central nervous system (CNS), and their uniquely protective role against subcellular mechanisms of degeneration induced by traumatic injury to the brain. We also discuss select studies on the therapeutic effects of n-3 PUFAs in vivo, and how omega-3 supplementation could potentially improve behavioral and cognitive outcomes in patients with TBI.

The Role of n-3 Fatty Acids in the Brain

The most important n-3 fatty acids for human health and nutrition are docosahexaenoic acid (DHA), eicosapentaenoic acid (EPA), and alpha-linolenic acid (ALA). Omega-3 fatty acids must be obtained from diet, and are highly enriched in algal oil, krill, and cold water fish. ^{20,21,190} Poultry and eggs also provide small quantities of EPA and DHA, ²² while nuts, soybean, canola, and flax seed oils are major dietary sources of ALA. ²³ Humans can also convert a limited amount of ALA into EPA and DHA, but synthesis of EPA and especially DHA from ALA is insufficient to supplant dietary intake. ²⁴

The human brain consists of 60% lipid by dry weight, and docosahexaenoic acid is one of the most abundant fatty acids found in the solid matter of the brain.²⁵ DHA is a primary structural component of the mammalian cerebral cortex, and comprises 50% of neuronal membrane phospholipids.²⁶ Omega-3 fatty acids are essential for maintaining membrane fluidity,²⁷ which in turn impacts neuronal cell adhesion, axon guidance, synapse maintenance, dendritic formation, and the speed of neurotransmission.²⁸⁻³⁰ DHA, for instance, is the longest and most unsaturated fatty acid found in biological membranes, with a structure that is tremendously flexible and therefore versatile.³¹ It is capable of undergoing rapid interconversions between multiple torsional states, and is unique in its ability to significantly alter membrane order and fluidity, phase behavior, elastic compressibility, ion permeability, and fusion. 32-39 Not surprisingly, then, this highly adaptable PUFA is particularly enriched in membranes which require rapid vesicle formation and release, such as rod outer segments in the retina and neuronal synapses. 40 The ability of DHA to readily undergo such complex yet minimal energy-requiring conformational changes is thought to be one of the main reasons why it is so abundant in the brain. 31 40

Due to its steric incompatibility with cholesterol and sphingomyelin, DHA also forms distinct lipid microdomains within the inner leaflet of the bilayer. 41 Such segregation is known to modulate the activity of various receptors, ion channels, G-proteins, and other membrane-bound proteins. 42 At physiological concentrations, for instance, DHA can potently and irreversibly inhibit ion flux through voltage-gated K⁺ channels in olfactory neurons.⁴³ Both DHA and EPA have additionally been shown to inhibit hippocampal neuron membrane excitability, whereas saturated and mono-unsaturated fatty acids do not. 44 Finally, a positive linear relationship also exists between the activity of the sodium-potassium pump (Na * K * ATPase) and the membrane concentration of DHA. 45 The mammalian brain has both the highest concentration of DHA as well as the highest activity rate of the sodium-potassium pump, and though the activity of Na⁺K⁺ATPase accounts for some 20% of the general basal metabolic rate, it accounts for approximately 60% of energy utilization in the brain.⁴⁵

An additional proposed reason for why omega-3 PUFAs are so essential to and enriched in the brain is their unique transformation into neuroprotective metabolites, which are critical in the defense against oxidative stress, tissue inflammation, and maintenance of synaptic integrity. For example, during tissue stress, both EPA and DHA are thought to be released from membrane phospholipids and converted into compounds called "resolvins," which actively promote resolution of inflammatory processes, such as via downregulation of NF- κ B and clearance of neutrophils. ^{49–51}

Nonmembrane bound (i.e., unesterified) DHA also regulates ion channel activity, as well as the expression of genes involved in control of signal transduction, synaptic plasticity, and cytoskeletal assembly. ^{23,29,52,53} In response to oxidative stress, free DHA may be used to synthesize neuroprotectin D1 (NPD1) via 15-lipoxygenase-1. NPD1, in turn, protects cells by upregulating anti-apoptotic proteins such as Bcl-2 and Bcl-xL, while downregulating pro-apoptotic proteins such as Bax and Bad. NPD1 synthesis can also be triggered in response to neurotrophins, ischemia, and reperfusion. ⁴⁷

Omega-3 polyunsaturated fatty acids are also ligands for retinoid X receptors (RXR) in the brain, which play a crucial role in neuronal growth and proliferation during fetal development, and in the morphological differentiation of catecholaminergic neurons. ^{54,55} RXR and related retinoic acid receptors are also highly expressed in the hippocampus, which may prove pertinent to understanding the functional role of omega-3s, particularly DHA, in the adult brain. ^{23,56}

Effects of n-3 Fatty Acid Deprivation

Chronic dietary deprivation of n-3 fatty acids in animals leads to:
1) decreased mean cell body size in neurons of the hippocampus, hypothalamus, and parietal cortex; 2) reduced complexity of dendritic arboritizations on cortical neurons; and 3) significant deficits in spatial learning and memory.^{57–61} In contrast, increased brain levels of DHA in adult mice enhances hippocampal neurogenesis as evidenced by an increased number of proliferating neurons, increased neurite outgrowth, and increased density of dendritic spines, all of which correlate with markedly improved performance in spatial learning tasks.⁶² In addition to serving as building blocks for membrane synthesis and modulating gene expression during neurogenesis, n-3 fatty acids are also involved in regulating

neurotransmitter receptor levels and activity. 58,59 Increased intake of n-3 PUFAs in a rat model of cerebral hypoperfusion, for instance, enriched the density of serotonergic and muscarinic acetylcholine (ACh) receptors in the dentate gyrus and CA3 regions of the hippocampus, and increased binding to muscarinic receptors. 63 Performance on memory tasks was also improved in these rats. 64

Rodents subjected to chronic omega-3 fatty acid deficiency also suffer from impaired attention and poor performance on shock avoidance, olfactory learning, exploratory, and flexibility behavior tasks. 65-68 Cholinergic pathways and ACh release are critical for arousal, attention, learning, and memory, 69-73 as is dopamine neurotransmission, which is also altered by dietary restriction of n-3 polyunsaturated fats. 74-77 Dopaminergic neurons in the mesolimbic system are critical for motivational behavior and emotional functions, while mesocortical dopaminergic neurons are key players in cognitive functions such as working memory. The vesicular storage pool of dopamine in both of these systems is depleted in n-3 PUFA-deficient animals, resulting in diminished dopamine release and impaired performance on cognitive tasks. To, 77, 79 The density of D2 dopamine receptors is also notably reduced in the frontal cortex of omega 3-deprived, aging rats.

Neurotherapeutic Effects of n-3 Fatty Acids in Vivo

Dietary supplementation has considerable influence on DHA content in the brain. R0,81 Though previous studies in mammals have focused heavily on chronic dietary deficiency, DHA has been shown to be neuroprotective in nondeficient animal models as well. In rats that had not been deprived of dietary fatty acids prior to traumatic axonal insult, for instance, DHA supplementation significantly ameliorated secondary mechanisms of injury and reduced the number of damaged axons.

Through mechanisms that are still incompletely understood, dietary supplementation with omega-3 fatty acids has been shown to decrease the production of reactive oxygen species (ROS) significantly and to improve cognitive function in vivo. 83,84 In ratbased models of ischemic injury, for instance, chronic administration of DHA reduced levels of lipid peroxidation byproducts and enhanced antioxidant activity in the brain. 83,85,86 Likewise, treatment at the onset of reperfusion has been shown to dramatically reduce infarction and increase scores on behavioral assessments.⁸⁷ Moreover, rats with traumatic spinal cord injuries have demonstrated significantly reduced inflammatory markers, along with increased neuron and oligodendrocyte survival and locomotor performance following a single dose of intravenous DHA postcompression or hemisection.^{88,89} When acute I.V. injection of DHA was followed with daily dietary supplementation, these therapeutic effects were sustained throughout the entire study duration of 6 weeks.⁸⁹ No significant improvement in outcome, however, was seen if intravenous DHA treatment was delayed until 3 hours post-injury, or if DHA was administered for 1 week through diet alone.

Recent reports have shown that omega-3 PUFAs may have a trophic effect on neurites as well, inducing more rapid and robust outgrowth of new fibers in animals fed a DHA-enriched diet after peripheral nerve transection. 90 Accelerated functional recovery and axonal regeneration secondary to upregulated n-3 PUFAs have also been observed in mice with acute traumatic injury to the sciatic nerve. 91 Similarly, in animal models of TBI, decreased levels of β -amyloid precursor protein, a marker of axonal injury, were observed after 1 month of dietary supplementation with DHA. 14,15 Decreased axonal injury counts and apoptotic markers as well as

improved memory have also been documented in rats with traumatic brain injuries when given prophylactic DHA for 30 days.⁸²

Polyunsaturated fatty acids in humans, particularly DHA, serve an essential role in nervous system development and are required for proper synaptogenesis, neural membrane synthesis, and the building of functionally critical circuits within the brain. DHA deficiency is associated with aging and neurodegenerative conditions such as Alzheimer's disease, while DHA consumption has been shown to improve performance on visuospatial learning and memory tasks in patients with age-related cognitive decline. There is additional evidence in humans that dietary supplementation with omega-3 fatty acids improves functional recovery in subarachnoid hemorrhage and stroke.

The impact of omega-3 PUFAs on prevention and recovery from stroke remains an intense area of inquiry, though the association between consumption of fish and fish oils and decreased risk of cardiovascular disease was first noted over 50 years ago. 98 Higher consumption of fish and n-3 fatty acids is correlated with a reduced risk of thrombotic stroke, 99,100 thus omega-3 PUFAs, in particular DHA, are now being investigated as putative "nutraceuticals" for treatment of cerebral ischemia. 101,102 Though the mechanisms by which omega-3s are protective against ischemic insult to the brain are not fully understood, neuroinflammation and programmed cell death are two well-known events underlying the pathophysiology of stroke. 103,104 DHA has previously been shown to be metabolized into neuroprotective mediators known as docosanoids, the most extensively studied of which is 10,17S-docosatriene, also referred to as the aforementioned NPD1. 48,105 NPD1 is a potent inhibitor of proinflammatory cytokine expression and apoptosis, as well as ischemia-reperfusion mediated infiltration by leukocytes. 46,101,106,107 DHA also has antioxidant activity and mitigates peroxidative damage of lipids and proteins in the brain.²³ In congruence with these findings, other studies have demonstrated that DHA can attenuate neuronal death and cognitive and locomotor impairments in animal models of ischemia-reperfusion injury to the brain. 108-110 Given that the same inflammatory, apoptotic, and oxidative stress mechanisms are implicated in traumatic injury to the brain, it can be reasonably hypothesized that accumulation of omega-3 fatty acids in the brain might also be neuroprotective in TBI.

Omega-3 Fatty Acids and the Pathogenesis of TBI

TBI results in a diffuse, progressive process of axonal destruction, demyelination, and neuronal cell death, not only at the site of impact but also in the surrounding parenchyma. ¹¹¹ Injury first occurs due to the direct physical forces associated with traumatic insult, but is then followed by a secondary wave of disruption in the subsequent hours and days due to inflammatory responses, excitotoxicity, and oxidative stress. ^{19,112,117,119} Omega-3 fatty acids mitigate the consequences of several key pathological pathways in TBI, such as mitochondrial malfunction, apoptotic cell death, glutamate-triggered excitotoxicity, and injury-induced oxidative stress and inflammation. ^{83,85,86,92,113,114,118} Omega-3 fatty acids may therefore play a critical role in the restoration of cellular energetics and repair of neuronal damage after TBI.

The production of pro-inflammatory prostaglandins is stimulated by and derived from the release of arachidonic acid (AA) secondary to disruption of neuronal cell membranes. Also released from neuronal membranes under pathological conditions, however, is DHA, which makes up 30% of the dry weight of the brain. Also Whereas arachidonic acid is rapidly

converted into potent inflammatory mediators such as prostaglandins, leukotrienes, and hydroxyeicosatetraenoic acids, DHA and its derivatives function in a neuroprotective capacity instead, antagonizing the pro-death signaling pathways initially triggered by AA. 92,113 The metabolism of two omega-3 fatty acids, EPA and DHA, specifically leads to the production of docosanoids (also known as "neuroprotectins") and resolvins, which not only inhibit the activation and migration of inflammatory cells, but upregulate anti-apoptotic cascades and expression of receptor families significantly involved in tissue repair as well. 114,187 These mechanisms, in turn, result in increased cell survivability and improved neurological outcome. 12,14–17

The nonspecific release of the excitotoxic neurotransmitter glutamate is another destructive event following acute traumatic injury in the brain. 117 Excess glutamate causes overactivation of N-methyl D-aspartate (NMDA) and calcium-permeable AMPA receptors, leading to massive influx of Ca2+ and the induction of both programmed and necrotic cell death via calcium-dependent proteases. 19,115-117 DHA has previously been shown to mitigate glutamate cytotoxicity and decrease Ca²⁺ influx in vitro, ¹¹⁸ and downregulates the expression of AMPA receptor subunits on the surface of cultured cells. 119 One of the most detrimental consequences of surplus intracellular Ca²⁺, however, is increased oxidative stress, a key contributor to the pathophysiologic changes that occur after TBI. Influx of excess Ca2+ into mitochondria leads to the formation of ROS, which directly damage DNA and proteins. The toxic accumulation of damaged DNA and oxidized proteins further induces programmed cell death. 117 ROS also initiate the process of lipid peroxidation on a catastrophic scale, which not only disrupts the integrity and function of neuronal membranes, but propagates further free radical formation secondary to the propensity of lipoperoxyl byproducts for attacking adjacent fatty acid chains. 113 Moreover, antioxidant defense mechanisms are relatively scarce in the human central nervous system, and the continued production of ROS via lipid peroxidation further depletes endogenous free radical scavengers that have already been overwhelmed. 120 Disruptions in cerebral blood flow likewise result in energy depletion and subsequent collapse of energy-dependent ion transport as well as intracellular Ca²⁺ overload. ¹⁹ 116 Hence oxidative stress is not limited to the ischemia/hypoxia stage; rather, a second course of oxidative damage (as well as inflammation) is incurred during the reperfusion phase of injury, resulting in additional microvascular damage, secondary ischemia, and neuronal cell death. 121 122

Clinical Considerations and Future Directions

TBI, with its intrinsic heterogeneity and prolonged secondary pathogenesis, remains a clinical challenge to manage. Clinical studies thus far have not yet identified an effective treatment strategy against secondary injury after traumatic insult to the brain has occurred. This is perhaps due to the fact that the majority of phase III prospective trials targeted single factors rather than multiple mechanisms of injury. MDA receptor antagonists, for example, were specifically aimed at reducing glutamate excitoxicity but failed to be of significant benefit in human trials. Likewise, administration of corticosteroids, well known for their anti-inflammatory effects, has shown no clear improvement in outcome or reduction in intracranial pressure, and one large study revealed that such compounds may in fact increase the risk of death after TBI. Other monotherapies that have been recently and clinically investigated for potential neuroprotection in

TBI include: cyclosporin A, progesterone, erythropoietin, and statins. 9,123

As most drugs aimed at limited pathways of injury have achieved little, if any, success in larger clinical trials, treatments with broader, pleiotropic effects are being increasingly explored. Multi-potential approaches such as dietary supplementation with omega-3 fatty acids, however, have primarily been investigated at only the pre-clinical stage. Two notable exceptions are progesterone and statins. The sex steroid progesterone, unlike corticosteroids, is thought to not only reduce cerebral edema but to also have neuroprotective effects as well, and has been positively correlated with improved functional outcomes at up to 6 months follow-up in two randomized, double-blind, placebo-controlled phase II trials. 128,129 Two multi-center, phase III clinical trials are now currently under way.

In addition to inhibiting cholesterol biosynthesis, 3-hydroxy-3-methylglutaryl coenzyme A (HMGCoA) reductase inhibitors, or "statins," exert favorable effects on multiple mechanisms of both acute and secondary injury. Larger phase II clinical trials specifically evaluating the use of rosuvastatin and artovastatin in the treatment of TBI have thus been planned. 123

Overall, multifunctional compounds such as HMGCoA reductase inhibitors have emerged as one of the most promising lines of defense against the deleterious effects of traumatic injury to the brain.

Like statins, n-3 PUFAs target multiple components of the secondary TBI injury cascade (Fig. 1) and, being an essential part of the human diet, are highly safe and well tolerated. 111,131,132 In contrast to statins and hormone-based therapies, omega-3s are readily taken up by the brain. Unlike hormones, they can be orally administered in addition to the parenteral route. 133,134 The U.S. Food and Drug Administration has furthermore confirmed the overall safety of fish oil, and both DHA and EPA at levels up to 3 g/day are generally recognized as safe. 135 The American Heart Association has furthermore established intakes of 1 g of EPA and DHA from fish or fish oils for patients with cardiovascular disease, and supplements of 2–4 g for subjects with high blood triglycerides. 136 Most clinical studies of DHA have employed a dose of 2–6 g/day, and no consistent adverse events have been observed in humans consuming from less than 1 up to 7.5g/day of DHA. 137

Potential harmful effects of n-3 PUFAs, however, have been described in the literature. Due to the established anti-thrombotic action of these compounds, for instance, they may increase the risk of hemorrhagic stroke, as suggested by a necropsy-based study of four cases in Greenland. 138 The authors warn, however, that the power of their analysis is weak given the limited sample size, and that their study may have been subject to inadvertent selection bias. 138 In addition, multiple clinical trials have shown that highdose fish oil consumption is safe, even in patients receiving other agents that may increase the risk of bleeding, such as aspirin and warfarin. 139-141 The overall clinical data suggests that DHA at doses up to 6 g/day does not have deleterious effects on platelet aggregation or other clotting parameters in normal individuals, and fish oil does not augment aspirin-induced inhibition of blood clotting. 137 Platelet function is, on the other hand, inhibited by DHA consumption in type 2 diabetics, but it is suggested that this may actually be of benefit to these individuals, especially when coupled with the other activities of DHA. 142 Nevertheless, it may be prudent to discontinue high-dose supplementation in the setting of an acute bleeding illness or in patients at high risk for hemorrhagic stroke or, as is frequently recommended with aspirin, warfarin, and clopidogrel, prior to planned invasive procedures with the highest risk for bleeding complications. 143-146

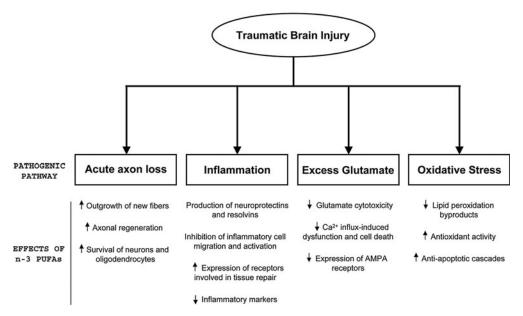


FIG. 1. Therapeutic impact of omega-3 fatty acids on different mechanisms of injury in TBI. Supplementation with n-3 polyunsaturated fatty acids improves cell survival, recovery, and functional outcome after traumatic insult via pleiotropic effects on a variety of pathways.

Despite the accumulation of evidence that DHA has protective antioxidant effects on cells, PUFAs nonetheless have high susceptibility to lipid peroxidation themselves by virtue of having acyl chains with several double bonds, 147 and lipoperoxidation byproducts, such as oxygenated α,β -unsaturated aldehydes that can be derived from oxidation of DHA, have a potential carcinogenic role. 148 Only one study has examined the formation of such compounds in vivo, and reported that plasma levels of 4-hydroxyhexenal (4-HHE), a lipoperoxyl specifically derived from DHA, were significantly increased in 12 men given 0.8-1.6 g/day DHA for 2 weeks. 149 The authors noted, on the other hand, that 4-HHE only represented 0.01% of plasma PUFAs in these men, suggesting that production of 4-HHE only took place to a very low extent. 149 Two studies have examined markers of oxidative stress in patients supplemented with 4 g/day and 6 g/day of DHA for 6 and 8 weeks, respectively, and found that lipid peroxidation was either unaffected or significantly lower in these individuals compared to placebo-treated controls. 150,151 Data on the potentially adverse effects of lipid oxidation following omega-3 fatty acid ingestion otherwise remain highly limited at this time.

High intake of fish oil via heavy consumption of fish may also increase the risk of exposure to environmental toxins and contaminants such as mercury and polychlorinated biphenyls. ^{152,153} The current body of evidence indicates that the benefits of fish intake generally outweigh the potential risks, except in a few selected species of fish. ¹⁵⁴ But the existing recommendations apply only to ingestion of fish rather than supplementation with purified fish oil, testing of select samples of which has revealed very low to negligible levels of mercury and other environmental toxins. ¹⁵⁵ Prescription preparations of fish oil additionally undergo even more rigorous purification and are subject to further regulatory processes and quality control in order to achieve FDA approval. ¹⁴³

Concern has also been raised regarding the possibility of immunosuppression by n-3 PUFAs due their anti-inflammatory effects. High doses of DHA (\geq 4.9 g/day), for instance, can suppress T-cell activation in humans and inhibit natural killer cell activity as well as decrease the number of total circulating white blood

cells.^{156–158} Although alterations in certain immune parameters may be of benefit in inflammatory diseases, they are of unknown consequence in healthy individuals. Intake of elevated levels of omega-3s may potentially incur a higher risk of infection, but no increased incidence or rate of infections has been reported thus far in the literature. Studies with sufficient statistical power to specifically examine this possibility, however, have not been conducted. ¹³⁷

A recent Cochrane Collaboration review concluded that fish oil supplementation may produce mild gastrointestinal discomfort but are otherwise well tolerated. The most common intolerance encountered to fish oil clinically is a "fishy" smell, aftertaste, and eructations. A practical solution to this problem, particularly in the setting of TBI, could be the use of intravenous as opposed to oral preparations of fish oil. In addition, vitamin E is frequently added to supplements in order to reduce rancidity, maintain freshness, and increase shelf life. 143

In summary, further clinical studies are warranted to examine closely both the potential benefits and adverse effects of omega-3 fatty acids in the acute phase of human TBI. To date, however, there have been no clinical trials examining nutritional intervention with omega-3s immediately after TBI. 18,19 There is a single report in which oral intake of n-3 and n-6 fatty acids for 90 days augmented immediate memory and attention scores in patients with mild cognitive impairment, as well as immediate and delayed memories in patients with organic brain lesions, including TBI. 161 It should be noted, however, that fewer than 10 patients with TBI were included in this study and all were required to be at least 5 years post-injury. Hence, supplementation was not initiated during the acute period but rather after a sustained, chronic history of neuropsychological decline. Furthermore, no physiological measures of neural activity were performed in these subjects, such as imaging via functional MRI. Finally, this study was performed prior to the validation and widespread use of biochemical markers in biological samples such as urine, serum, and CSF to both monitor and quantify primary and evolving damage in TBI, such as the highly brain-specific protein S100B. 162-165 Serum levels of S100B have recently been shown to

predict CT findings and clinical outcomes in mild traumatic head injury, ¹⁶⁶ and are increasingly utilized to help identify, for example, patients who may benefit from early surgical intervention after TBI. ¹⁶⁴ Future studies on the neuroprotective potential of n-3 fatty acids for treatment of TBI could therefore examine whether sustained omega-3 administration can reduce levels of biomarkers of structural damage and inflammation, enhance regional brain activity, and improve cognitive function in post-traumatic patients over time.

TBI is an intrinsically multifaceted disease and therefore requires a combinatorial approach to its management. Nutritional interventions targeting key pathological factors in the acute phase will differ from those directed toward the subacute and chronic phase of TBI. Orally administered, omega-3 fatty acids may take days to weeks to get incorporated into cellular membranes to demonstrate the potential benefits. Therefore, intravenous administration of omega-3 fatty acids could be a more suitable intervention to study the immediate potential benefits after TBI, while sustained oral administration may enhance the repair and recovery mechanisms after TBI.

Conclusion

Omega-3 fatty acids restore cellular energetics, reduce oxidative stress and inflammation, repair cellular damage, and mitigate the activation of apoptotic processes after TBI. Simultaneously affecting these well-elucidated key pathological mechanisms associated with TBI, well tolerated, and easy to administer, nutritional interventions using omega-3 fatty acids present a unique advantage and opportunity. Further clinical studies are warranted to examine the potential benefits closely, as well as any drawbacks, of omega-3 fatty acids as an integral component of multidisciplinary treatment to lessen both the primary and secondary effects of TBI.

Author Disclosure Statement

No competing financial interests exist.

References

- Warden D. (2006). Military TBI during the Iraq and Afghanistan wars. J Head Trauma Rehabil 21, 398–402.
- Langlois JA, Rutland-Brown W, and Wald MM. (2006). The epidemiology and impact of traumatic brain injury: A brief overview. J Head Trauma Rehabil 21, 375–378.
- Schroeder T, Irish C, Annest JL, Haileyesus T, Sarmiento K, and Mitchko J. (2007). Nonfatal traumatic brain injuries from sports and recreation activities—United States, 2001–2005. Morbidity Mortality Weekly Rev Center for Disease Control and Prevention, p 733–737.
- Thurman DJ, Alverson C, Dunn KA, Guerrero J, and Sniezek JE. (1999). Traumatic brain injury in the United States: A public health perspective. J Head Trauma Rehabil 14, 602–615.
- Solomon GS, Ott SD, and Lovell MR. (2011). Long-term neurocognitive dysfunction in sports: What is the evidence? Clin Sports Med 30, 165–177.
- Omalu BI, DeKosky ST, Hamilton RL, et al. (2006). Chronic traumatic encephalopathy in a national football league player: Part II. Neurosurgery 59, 1086–1092; discussion 92–93.
- Guskiewicz KM, Marshall SW, Bailes J, et al. (2007). Recurrent concussion and risk of depression in retired professional football players. Med Sci Sports Exerc 39, 903–909.
- Omalu BI, Bailes J, Hammers JL, and Fitzsimmons RP. (2010). Chronic traumatic encephalopathy, suicides and parasuicides in professional American athletes: The role of the forensic pathologist. Am J Forensic Med Pathol 31, 130–132.
- 9. Xiong Y, Mahmood A, and Chopp M. (2009). Emerging treatments for traumatic brain injury. Expert Opin Emerg Drugs 14, 67–84.

 Wu A, Ying Z, and Gomez-Pinilla F. (2008). Docosahexaenoic acid dietary supplementation enhances the effects of exercise on synaptic plasticity and cognition. Neuroscience 155, 751–759.

- Lang-Lazdunski L, Blondeau N, Jarretou G, Lazdunski M, and Heurteaux C. (2003). Linolenic acid prevents neuronal cell death and paraplegia after transient spinal cord ischemia in rats. J Vasc Surg 38, 564–575.
- Wu A, Ying Z, and Gomez-Pinilla F. (2004). Dietary omega-3 fatty acids normalize BDNF levels, reduce oxidative damage, and counteract learning disability after traumatic brain injury in rats. J Neurotrauma 21, 1457–1467.
- Wu A, Ying Z, and Gomez-Pinilla F. (2011). The salutary effects of DHA dietary supplementation on cognition, neuroplasticity, and membrane homeostasis after brain trauma. J Neurotrauma 28, 2113– 2122
- Mills JD, Bailes JE, Sedney CL, Hutchins H, and Sears B. (2011).
 Omega-3 fatty acid supplementation and reduction of traumatic axonal injury in a rodent head injury model. J Neurosurg 114, 77–84.
- Mills JD, Hadley K, and Bailes JE. (2011). Dietary supplementation with the omega-3 fatty acid docosahexaenoic acid in traumatic brain injury. Neurosurgery 68, 474–481; discussion 481.
- Prins M. (2008). Diet, ketones, and neurotrauma. Epilepsia 49, 111–113.
- Shin SS, and Dixon CE. (2011). Oral fish oil restores striatal dopamine release after traumatic brain injury. Neurosci Lett 496, 168–171.
- Petraglia AL, Winkler EA, and Bailes JE. (2011). Stuck at the bench: Potential natural neuroprotective compounds for concussion. Surg Neurol Int 2, 146.
- National Research Council (2011). Nutrition and Traumatic Brain Injury: Improving Acute and Subacute Outcomes in Military Personnel. Erdman J, Oria M, Pillsbury L (eds.) Washington, D.C.: The National Academies Press, 2011.
- Kris-Etherton PM, Taylor DS, Yu-Poth S, et al. (2000). Polyunsaturated fatty acids in the food chain in the United States. Am J Clin Nutr 71, 179S–88S.
- Raper NR, Cronin FJ, and Exler J. (1992). Omega-3 fatty acid content of the US food supply. J Am Coll Nutr 11, 304–308.
- 22. Innis SM. (2003). Perinatal biochemistry and physiology of longchain polyunsaturated fatty acids. J Pediatr 143, S1–8.
- Innis SM. (2007). Dietary (n-3) fatty acids and brain development. J Nutr 137, 855–859.
- 24. Goyens PL, Spilker ME, Zock PL, Katan MB, and Mensink RP. (2006). Conversion of alpha-linolenic acid in humans is influenced by the absolute amounts of alpha-linolenic acid and linoleic acid in the diet and not by their ratio. Am J Clin Nutr 84, 44–53.
- Crawford MA. (1993). The role of essential fatty acids in neural development: Implications for perinatal nutrition. Am J Clin Nutr 57, 703S–09S; discussion 09S–10S.
- Singh M. (2005). Essential fatty acids, DHA and human brain. Indian J Pediatr 72, 239–242.
- Valentine RC, and Valentine DL. (2004). Omega-3 fatty acids in cellular membranes: A unified concept. Prog Lipid Res 43, 383–402.
- Hering H, Lin CC, and Sheng M. (2003). Lipid rafts in the maintenance of synapses, dendritic spines, and surface AMPA receptor stability. J Neurosci 23, 3262–3271.
- 29. Innis SM. (2008). Dietary omega 3 fatty acids and the developing brain. Brain Res 1237, 35–43.
- Tsui-Pierchala BA, Encinas M, Milbrandt J, and Johnson EM, Jr. (2002). Lipid rafts in neuronal signaling and function. Trends Neurosci 25, 412–417.
- Stillwell W, and Wassall SR. (2003). Docosahexaenoic acid: Membrane properties of a unique fatty acid. Chem Phys Lipids 126, 1–27.
- Armstrong VT, Brzustowicz MR, Wassall SR, Jenski LJ, and Stillwell W. (2003). Rapid flip-flop in polyunsaturated (docosahexaenoate) phospholipid membranes. Arch Biochem Biophys 414, 74–82.
- Chapkin RS, Wang N, Fan YY, Lupton JR, and Prior IA. (2008).
 Docosahexaenoic acid alters the size and distribution of cell surface microdomains. Biochim Biophys Acta 1778, 466–471.
- 34. Ehringer W, Belcher D, Wassall SR, and Stillwell W. (1990). A comparison of the effects of linolenic (18:3 omega 3) and docosahexaenoic (22:6 omega 3) acids on phospholipid bilayers. Chem Phys Lipids 54, 79–88.

- Huster D, Arnold K, and Gawrisch K. (1998). Influence of docosahexaenoic acid and cholesterol on lateral lipid organization in phospholipid mixtures. Biochemistry 37, 17299–17308.
- Kafrawy O, Zerouga M, Stillwell W, and Jenski LJ. (1998). Docosahexaenoic acid in phosphatidylcholine mediates cytotoxicity more effectively than other omega-3 and omega-6 fatty acids. Cancer Lett 132, 23–29.
- Salem N, Jr., and Niebylski CD. (1995). The nervous system has an absolute molecular species requirement for proper function. Mol Membr Biol 12, 131–134.
- Smaby JM, Momsen MM, Brockman HL, and Brown RE. (1997).
 Phosphatidylcholine acyl unsaturation modulates the decrease in interfacial elasticity induced by cholesterol. Biophys J 73, 1492– 1505.
- Wassall SR, and Stillwell W. (2008). Docosahexaenoic acid domains: The ultimate non-raft membrane domain. Chem Phys Lipids 153, 57–63
- Stillwell W, Shaikh SR, Zerouga M, Siddiqui R, and Wassall SR. (2005). Docosahexaenoic acid affects cell signaling by altering lipid rafts. Reprod Nutr Dev 45, 559–579.
- Shaikh SR, Cherezov V, Caffrey M, Stillwell W, and Wassall SR. (2003). Interaction of cholesterol with a docosahexaenoic acidcontaining phosphatidylethanolamine: trigger for microdomain/raft formation? Biochemistry 42, 12028–12037.
- Wurtman RJ. (2008). Synapse formation and cognitive brain development: Effect of docosahexaenoic acid and other dietary constituents. Metabolism 57, S6–10.
- Seebungkert B, and Lynch JW. (2002). Effects of polyunsaturated fatty acids on voltage-gated K+ and Na+ channels in rat olfactory receptor neurons. Eur J Neurosci 16, 2085–2094.
- Xiao Y, and Li X. (1999). Polyunsaturated fatty acids modify mouse hippocampal neuronal excitability during excitotoxic or convulsant stimulation. Brain Res 846, 112–121.
- Turner N, Else PL, and Hulbert AJ. (2003). Docosahexaenoic acid (DHA) content of membranes determines molecular activity of the sodium pump: Implications for disease states and metabolism. Naturwissenschaften 90, 521–523.
- Bazan NG. (2005). Neuroprotectin D1 (NPD1): A DHA-derived mediator that protects brain and retina against cell injury-induced oxidative stress. Brain Pathol 15, 159–166.
- Bazan NG, Musto AE, and Knott EJ. (2011). Endogenous signaling by omega-3 docosahexaenoic acid-derived mediators sustains homeostatic synaptic and circuitry integrity. Mol Neurobiol 44, 216– 222.
- 48. Niemoller TD, Stark DT, and Bazan NG. (2009). Omega-3 fatty acid docosahexaenoic acid is the precursor of neuroprotectin D1 in the nervous system. World Rev Nutr Diet 99, 46–54.
- 49. Weylandt KH, Chiu CY, Gomolka B, Waechter SF, and Wiedenmann B. (2012). Omega-3 fatty acids and their lipid mediators: Towards an understanding of resolvin and protectin formation. Prostaglandins Other Lipid Mediat 97, 73–82.
- Tassoni D, Kaur G, Weisinger RS, and Sinclair AJ. (2008). The role of eicosanoids in the brain. Asia Pac J Clin Nutr 17, 220–228.
- Im DS. (2012). Omega-3 fatty acids in anti-inflammation (proresolution) and GPCRs. Prog Lipid Res 51, 232–237.
- Horrocks LA, and Farooqui AA. (2004). Docosahexaenoic acid in the diet: Its importance in maintenance and restoration of neural membrane function. Prostaglandins Leukot Essent Fatty Acids 70, 361– 372.
- 53. Kitajka K, Puskas LG, Zvara A, et al. (2002). The role of n-3 polyunsaturated fatty acids in brain: Modulation of rat brain gene expression by dietary n-3 fatty acids. Proc Natl Acad Sci USA 99, 2619–2624
- Lengqvist J, Mata De Urquiza A, Bergman AC, et al. (2004).
 Polyunsaturated fatty acids including docosahexaenoic and arachidonic acid bind to the retinoid X receptor alpha ligand-binding domain. Mol Cell Proteomics 3, 692–703.
- 55. Rioux L, and Arnold SE. (2005). The expression of retinoic acid receptor alpha is increased in the granule cells of the dentate gyrus in schizophrenia. Psychiatry Res 133, 13–21.
- 56. Lane MA, and Bailey SJ. (2005). Role of retinoid signalling in the adult brain. Prog Neurobiol 75, 275–293.
- Ahmad A, Moriguchi T, and Salem N. (2002). Decrease in neuron size in docosahexaenoic acid-deficient brain. Pediatr Neurol 26, 210– 218.

- Aid S, Vancassel S, Poumes-Ballihaut C, Chalon S, Guesnet P, and Lavialle M. (2003). Effect of a diet-induced n-3 PUFA depletion on cholinergic parameters in the rat hippocampus. J Lipid Res 44, 1545– 1551.
- Chalon S. (2006). Omega-3 fatty acids and monoamine neurotransmission. Prostaglandins Leukot Essent Fatty Acids 75, 259–269.
- Innis SM, Gilley J, and Werker J. (2001). Are human milk long-chain polyunsaturated fatty acids related to visual and neural development in breast-fed term infants? J Pediatr 139, 532–538.
- Wainwright RE, Bulman-Fleming MB, Lévesque S, Mutsaers L, and McCutcheon D. (1998). A saturated-fat diet during development alters dendritic growth in mouse brain. Nutr Neurosci 1, 49–58.
- 62. He C, Qu X, Cui L, Wang J, and Kang JX. (2009). Improved spatial learning performance of fat-1 mice is associated with enhanced neurogenesis and neuritogenesis by docosahexaenoic acid. Proc Natl Acad Sci USA 106, 11370–11375.
- 63. Farkas E, de Wilde MC, Kiliaan AJ, Meijer J, Keijser JN, and Luiten PG. (2002). Dietary long chain PUFAs differentially affect hippocampal muscarinic 1 and serotonergic 1A receptors in experimental cerebral hypoperfusion. Brain Res 954, 32–41.
- Farkas E, de Wilde MC, Kiliaan AJ, and Luiten PG. (2002). Systemic
 effects of dietary n-3 PUFA supplementation accompany changes of
 CNS parameters in cerebral hypoperfusion. Ann NY Acad Sci 977,
 77–86
- Catalan J, Moriguchi T, Slotnick B, Murthy M, Greiner RS, and Salem N, Jr. (2002). Cognitive deficits in docosahexaenoic aciddeficient rats. Behav Neurosci 116, 1022–1031.
- 66. Bourre JM, Francois M, Youyou A, et al. (1989). The effects of dietary alpha-linolenic acid on the composition of nerve membranes, enzymatic activity, amplitude of electrophysiological parameters, resistance to poisons and performance of learning tasks in rats. J Nutr 119, 1880–1892.
- Carrie I, Clement M, de Javel D, Frances H, and Bourre JM. (2000).
 Phospholipid supplementation reverses behavioral and biochemical alterations induced by n-3 polyunsaturated fatty acid deficiency in mice. J Lipid Res 41, 473–480.
- Yamamoto N, Hashimoto A, Takemoto Y, et al. (1988). Effect of the dietary alpha-linolenate/linoleate balance on lipid compositions and learning ability of rats. II. Discrimination process, extinction process, and glycolipid compositions. J Lipid Res 29, 1013–1021.
- Blokland A. (1995). Acetylcholine: A neurotransmitter for learning and memory? Brain Res Brain Res Rev 21, 285–300.
- Everitt BJ, and Robbins TW. (1997). Central cholinergic systems and cognition. Annu Rev Psychol 48, 649–684.
- Fadda F, Cocco S, and Stancampiano R. (2000). Hippocampal acetylcholine release correlates with spatial learning performance in freely moving rats. Neuroreport 11, 2265–2269.
- Himmelheber AM, Sarter M, and Bruno JP. (2000). Increases in cortical acetylcholine release during sustained attention performance in rats. Brain Res Cogn Brain Res 9, 313–325.
- Passetti F, Dalley JW, O'Connell MT, Everitt BJ, and Robbins TW. (2000). Increased acetylcholine release in the rat medial prefrontal cortex during performance of a visual attentional task. Eur J Neurosci 12, 3051–3058.
- Chalon S, Vancassel S, Zimmer L, Guilloteau D, and Durand G. (2001). Polyunsaturated fatty acids and cerebral function: Focus on monoaminergic neurotransmission. Lipids 36, 937–944.
- Delion S, Chalon S, Guilloteau D, Besnard JC, and Durand G. (1996). alpha-Linolenic acid dietary deficiency alters age-related changes of dopaminergic and serotoninergic neurotransmission in the rat frontal cortex. J Neurochem 66, 1582–1591.
- Zimmer L, Delion-Vancassel S, Durand G, et al. (2000). Modification of dopamine neurotransmission in the nucleus accumbens of rats deficient in n-3 polyunsaturated fatty acids. J Lipid Res 41, 32–40.
- Zimmer L, Vancassel S, Cantagrel S, et al. (2002). The dopamine mesocorticolimbic pathway is affected by deficiency in n-3 polyunsaturated fatty acids. Am J Clin Nutr 75, 662–667.
- Mathe JM, Nomikos GG, Blakeman KH, and Svensson TH. (1999).
 Differential actions of dizocilpine (MK-801) on the mesolimbic and mesocortical dopamine systems: Role of neuronal activity. Neuropharmacology 38, 121–128.
- Zimmer L, Delpal S, Guilloteau D, Aioun J, Durand G, and Chalon S. (2000). Chronic n-3 polyunsaturated fatty acid deficiency alters dopamine vesicle density in the rat frontal cortex. Neurosci Lett 284, 25–28.

 Brenna JT, and Diau GY. (2007). The influence of dietary docosahexaenoic acid and arachidonic acid on central nervous system polyunsaturated fatty acid composition. Prostaglandins Leukot Essent Fatty Acids 77, 247–250.

- Diau GY, Hsieh AT, Sarkadi-Nagy EA, Wijendran V, Nathanielsz PW, and Brenna JT. (2005). The influence of long chain polyunsaturate supplementation on docosahexaenoic acid and arachidonic acid in baboon neonate central nervous system. BMC Med 3:11.
- Bailes JE, and Mills JD. (2010). Docosahexaenoic acid reduces traumatic axonal injury in a rodent head injury model. J Neurotrauma 27, 1617–1624.
- Pan HC, Kao TK, Ou YC, et al. (2009). Protective effect of docosahexaenoic acid against brain injury in ischemic rats. J Nutr Biochem 20, 715–725.
- 84. Suchy J, Chan A, and Shea TB. (2009). Dietary supplementation with a combination of alpha-lipoic acid, acetyl-L-carnitine, glycerophosphocoline, docosahexaenoic acid, and phosphatidylserine reduces oxidative damage to murine brain and improves cognitive performance. Nutr Res 29, 70–74.
- 85. Choi-Kwon S, Park KA, Lee HJ, et al. (2004). Temporal changes in cerebral antioxidant enzyme activities after ischemia and reperfusion in a rat focal brain ischemia model: Effect of dietary fish oil. Brain Res Dev Brain Res 152, 11–18.
- Glozman S, Green P, and Yavin E. (1998). Intraamniotic ethyl docosahexaenoate administration protects fetal rat brain from ischemic stress. J Neurochem 70, 2484

 –2491.
- Belayev L, Marcheselli VL, Khoutorova L, et al. (2005). Docosahexaenoic acid complexed to albumin elicits high-grade ischemic neuroprotection. Stroke 36, 118–123.
- Huang WL, King VR, Curran OE, et al. (2007). A combination of intravenous and dietary docosahexaenoic acid significantly improves outcome after spinal cord injury. Brain 130, 3004–3019.
- King VR, Huang WL, Dyall SC, Curran OE, Priestley JV, and Michael-Titus AT. (2006). Omega-3 fatty acids improve recovery, whereas omega-6 fatty acids worsen outcome, after spinal cord injury in the adult rat. J Neurosci 26, 4672–4680.
- Effect of docosahexaenoic acid, an omega-3 polyunsaturated fatty acid, in a mouse facial nerve injury model. British Pharmacological Society 75th Anniversary Meeting; 2006. Proceedings of the British Pharmacological Society.
- Gladman SJ, Huang W, Lim SN, et al. (2012). Improved outcome after peripheral nerve injury in mice with increased levels of endogenous omega-3 polyunsaturated fatty acids. J Neurosci 32, 563– 571
- Bazan NG, Molina MF, and Gordon WC. (2011). Docosahexaenoic acid signalolipidomics in nutrition: Significance in aging, neuroinflammation, macular degeneration, Alzheimer's, and other neurodegenerative diseases. Annu Rev Nutr 31, 321–351.
- Yurko-Mauro K, McCarthy D, Rom D, et al. (2010). Beneficial effects of docosahexaenoic acid on cognition in age-related cognitive decline. Alzheimers Dement 6, 456–464.
- Bistrian BR, Askew W, Erdman JW, Jr., and Oria MP. (2011). Nutrition and traumatic brain injury: A perspective from the Institute of Medicine report. JPEN J Parenter Enteral Nutr 35, 556–9.
- 95. Garbagnati F, Cairella G, De Martino A, et al. (2009). Is antioxidant and n-3 supplementation able to improve functional status in poststroke patients? Results from the Nutristroke Trial. Cerebrovasc Dis 27, 375–383.
- Poppitt SD, Howe CA, Lithander FE, et al. (2009). Effects of moderate-dose omega-3 fish oil on cardiovascular risk factors and mood after ischemic stroke: A randomized, controlled trial. Stroke 40, 3485–3492.
- Yoneda H, Shirao S, Kurokawa T, Fujisawa H, Kato S, and Suzuki M. (2008). Does eicosapentaenoic acid (EPA) inhibit cerebral vasospasm in patients after aneurysmal subarachnoid hemorrhage? Acta Neurol Scand 118, 54–59.
- 98. Skerrett PJ, and Hennekens CH. (2003). Consumption of fish and fish oils and decreased risk of stroke. Prev Cardiol 6, 38–41.
- He K, Rimm EB, Merchant A, et al. (2002). Fish consumption and risk of stroke in men. JAMA 288, 3130–3136.
- Iso H, Rexrode KM, Stampfer MJ, et al. (2001). Intake of fish and omega-3 fatty acids and risk of stroke in women. JAMA 285, 304– 312.
- Lalancette-Hebert M, Julien C, Cordeau P, et al. (2011). Accumulation of dietary docosahexaenoic acid in the brain attenuates acute

- immune response and development of postischemic neuronal damage. Stroke 42, 2903–2909.
- 102. Mozaffarian D, Longstreth WT, Jr., Lemaitre RN, et al. (2005). Fish consumption and stroke risk in elderly individuals: The cardiovascular health study. Arch Intern Med 165, 200–206.
- Dirnagl U, Iadecola C, and Moskowitz MA. (1999). Pathobiology of ischaemic stroke: An integrated view. Trends Neurosci 22, 391–397.
- 104. Moskowitz MA, Lo EH, and Iadecola C. (2010). The science of stroke: Mechanisms in search of treatments. Neuron 67, 181–198.
- 105. Mukherjee PK, Marcheselli VL, Serhan CN, and Bazan NG. (2004). Neuroprotectin D1: A docosahexaenoic acid-derived docosatriene protects human retinal pigment epithelial cells from oxidative stress. Proc Natl Acad Sci USA 101, 8491–8496.
- 106. Bazan NG. (2009). Neuroprotectin D1-mediated anti-inflammatory and survival signaling in stroke, retinal degenerations, and Alzheimer's disease. J Lipid Res 50, S400–405.
- Marcheselli VL, Hong S, Lukiw WJ, et al. (2003). Novel docosanoids inhibit brain ischemia-reperfusion-mediated leukocyte infiltration and pro-inflammatory gene expression. J Biol Chem 278, 43807–43817.
- Cao DH, Xu JF, Xue RH, Zheng WF, and Liu ZL. (2004). Protective effect of chronic ethyl docosahexaenoate administration on brain injury in ischemic gerbils. Pharmacol Biochem Behav 79, 651–659
- 109. Green P, Glozman S, Weiner L, and Yavin E. (2001). Enhanced free radical scavenging and decreased lipid peroxidation in the rat fetal brain after treatment with ethyl docosahexaenoate. Biochim Biophys Acta 1532, 203–212.
- 110. Okada M, Amamoto T, Tomonaga M, et al. (1996). The chronic administration of docosahexaenoic acid reduces the spatial cognitive deficit following transient forebrain ischemia in rats. Neuroscience 71, 17–25.
- Farooqui AA. (2010). Potential neuroprotective strategies for traumatic brain injury. In: Neurochemical Aspects of Neurotraumatic and Neurodegenerative Diseases. 1st ed: Springer, pp. 219–248.
- Dyall SC, and Michael-Titus AT. (2008). Neurological benefits of omega-3 fatty acids. Neuromolecular Med 10, 219–235.
- Farooqui AA, Horrocks LA, and Farooqui T. (2007). Modulation of inflammation in brain: A matter of fat. J Neurochem 101, 577–599.
- Bazan NG. (2006). Cell survival matters: Docosahexaenoic acid signaling, neuroprotection and photoreceptors. Trends Neurosci 29, 263–271.
- Artal-Sanz M, and Tavernarakis N. (2005). Proteolytic mechanisms in necrotic cell death and neurodegeneration. FEBS Lett 579, 3287– 3296.
- Choi DW, and Rothman SM. (1990). The role of glutamate neurotoxicity in hypoxic-ischemic neuronal death. Annu Rev Neurosci 13, 171–182.
- Obrenovitch TP, and Urenjak J. (1997). Is high extracellular glutamate the key to excitotoxicity in traumatic brain injury? J Neurotrauma 14, 677–698.
- Wang X, Zhao X, Mao ZY, Wang XM, and Liu ZL. (2003). Neuroprotective effect of docosahexaenoic acid on glutamate-induced cytotoxicity in rat hippocampal cultures. Neuroreport 14, 2457–2461.
- 119. Menard C, Patenaude C, Gagne AM, and Massicotte G. (2009). AMPA receptor-mediated cell death is reduced by docosahexaenoic acid but not by eicosapentaenoic acid in area CA1 of hippocampal slice cultures. J Neurosci Res 87, 876–886.
- Pratico D, Reiss P, Tang LX, Sung S, Rokach J, and McIntosh TK. (2002). Local and systemic increase in lipid peroxidation after moderate experimental traumatic brain injury. J Neurochem 80, 894– 898.
- Hall ED, and Springer JE. (2004). Neuroprotection and acute spinal cord injury: A reappraisal. NeuroRx 1, 80–100.
- 122. Xu W, Chi L, Xu R, et al. (2005). Increased production of reactive oxygen species contributes to motor neuron death in a compression mouse model of spinal cord injury. Spinal Cord 43, 204–213.
- Loane DJ, and Faden AI. (2010). Neuroprotection for traumatic brain injury: Translational challenges and emerging therapeutic strategies. Trends Pharmacol Sci 31, 596–604.
- 124. Ikonomidou C, and Turski L. (2002). Why did NMDA receptor antagonists fail clinical trials for stroke and traumatic brain injury? Lancet Neurol 1, 383–386.
- 125. Maas AI, Murray G, Henney H, 3rd, et al. (2006). Efficacy and safety of dexanabinol in severe traumatic brain injury: Results of a phase III

- randomised, placebo-controlled, clinical trial. Lancet Neurol 5, 38-45.
- 126. Narayan RK, Michel ME, Ansell B, et al. (2002). Clinical trials in head injury. J Neurotrauma 19, 503–557.
- Alderson P, and Roberts I. (2005). Corticosteroids for acute traumatic brain injury. Cochrane Database Syst Rev(1):CD000196.
- 128. Wright DW, Kellermann AL, Hertzberg VS, et al. (2007). ProTECT: A randomized clinical trial of progesterone for acute traumatic brain injury. Ann Emerg Med 49, 391–402, 02 e1–2.
- 129. Xiao G, Wei J, Yan W, Wang W, and Lu Z. (2008). Improved outcomes from the administration of progesterone for patients with acute severe traumatic brain injury: A randomized controlled trial. Crit Care 12, R61.
- 130. Wible EF, and Laskowitz DT. (2010). Statins in traumatic brain injury. Neurotherapeutics 7, 62–73.
- 131. Davidson MH, Stein EA, Bays HE, et al. (2007). Efficacy and tolerability of adding prescription omega-3 fatty acids 4 g/d to simvastatin 40 mg/d in hypertriglyceridemic patients: An 8-week, randomized, double-blind, placebo-controlled study. Clin Ther 29, 1354–1367.
- 132. Wohl DA, Tien HC, Busby M, et al. (2005). Randomized study of the safety and efficacy of fish oil (omega-3 fatty acid) supplementation with dietary and exercise counseling for the treatment of anti-retroviral therapy-associated hypertriglyceridemia. Clin Infect Dis 41, 1498–1504.
- Ott J, Hiesgen C, and Mayer K. (2011). Lipids in critical care medicine. Prostaglandins Leukot Essent Fatty Acids 85, 267–273.
- 134. Rapoport SI, Rao JS, and Igarashi M. (2007). Brain metabolism of nutritionally essential polyunsaturated fatty acids depends on both the diet and the liver. Prostaglandins Leukot Essent Fatty Acids 77, 251–261.
- 135. Substances affirmed as generally recognized as safe: Menhaden oil. *Final Rule*: Federal Registry, 1997, 30751–30757.
- Kris-Etherton PM, Harris WS, and Appel LJ. (2003). Omega-3 fatty acids and cardiovascular disease: New recommendations from the American Heart Association. Arterioscler Thromb Vasc Biol 23, 151–152.
- 137. Lien EL. (2009). Toxicology and safety of DHA. Prostaglandins Leukot Essent Fatty Acids 81, 125–132.
- Pedersen HS, Mulvad G, Seidelin KN, Malcom GT, and Boudreau DA. (1999). N-3 fatty acids as a risk factor for haemorrhagic stroke. Lancet 353, 812–813.
- 139. Bender NK, Kraynak MA, Chiquette E, Linn WD, Clark GM, and Bussey HI. (1998). Effects of marine fish oils on the anticoagulation status of patients receiving chronic warfarin therapy. J Thromb Thrombolysis 5, 257–261.
- 140. Eritsland J, Arnesen H, Gronseth K, Fjeld NB, and Abdelnoor M. (1996). Effect of dietary supplementation with n-3 fatty acids on coronary artery bypass graft patency. Am J Cardiol 77, 31–36.
- 141. Leaf A, Jorgensen MB, Jacobs AK, et al. (1994). Do fish oils prevent restenosis after coronary angioplasty? Circulation 90, 2248–2257.
- 142. Woodman RJ, Mori TA, Burke V, et al. (2003). Effects of purified eicosapentaenoic acid and docosahexaenoic acid on platelet, fibrinolytic and vascular function in hypertensive type 2 diabetic patients. Atherosclerosis 166, 85–93.
- Bays HE. (2007). Safety considerations with omega-3 fatty acid therapy. Am J Cardiol 99, 35C–43C.
- 144. Calo L, Bianconi L, Colivicchi F, et al. (2005). N-3 Fatty acids for the prevention of atrial fibrillation after coronary artery bypass surgery: A randomized, controlled trial. J Am Coll Cardiol 45, 1723– 1728.
- 145. Covington MB. (2004). Omega-3 fatty acids. Am Fam Physician 70, 133-140
- 146. Kris-Etherton PM, Harris WS, and Appel LJ. (2003). Fish consumption, fish oil, omega-3 fatty acids, and cardiovascular disease. Arterioscler Thromb Vasc Biol 23, e20–30.
- Cosgrove JP, Church DF, and Pryor WA. (1987). The kinetics of the autoxidation of polyunsaturated fatty acids. Lipids 22, 299–304.
- 148. Guillen MD, and Goicoechea E. (2008). Toxic oxygenated alpha, beta-unsaturated aldehydes and their study in foods: A review. Crit Rev Food Sci Nutr 48, 119–136.
- 149. Calzada C, Colas R, Guillot N, et al. (2010). Subgram daily supplementation with docosahexaenoic acid protects low-density lipoproteins from oxidation in healthy men. Atherosclerosis 208, 467– 472

- 150. Mori TA, Woodman RJ, Burke V, Puddey IB, Croft KD, and Beilin LJ. (2003). Effect of eicosapentaenoic acid and docosahexaenoic acid on oxidative stress and inflammatory markers in treated-hypertensive type 2 diabetic subjects. Free Radic Biol Med 35, 772–781
- 151. Wu WH, Lu SC, Wang TF, Jou HJ, and Wang TA. (2006). Effects of docosahexaenoic acid supplementation on blood lipids, estrogen metabolism, and in vivo oxidative stress in postmenopausal vegetarian women. Eur J Clin Nutr 60, 386–92.
- 152. Arisawa K, Matsumura T, Tohyama C, et al. (2003). Fish intake, plasma omega-3 polyunsaturated fatty acids, and polychlorinated dibenzo-purians and co-planar polychlorinated biphenyls in the blood of the Japanese population. Int Arch Occup Environ Health 76, 205–215.
- 153. Mulvad G, Pedersen HS, Hansen JC, et al. (1996). The Inuit diet. Fatty acids and antioxidants, their role in ischemic heart disease, and exposure to organochlorines and heavy metals. An international study. Arctic Med Res 55, 20–24.
- 154. Mozaffarian D, and Rimm EB. (2006). Fish intake, contaminants, and human health: Evaluating the risks and the benefits. JAMA 296, 1885–1899.
- 155. Foran SE, Flood JG, and Lewandrowski KB. (2003). Measurement of mercury levels in concentrated over-the-counter fish oil preparations: Is fish oil healthier than fish? Arch Pathol Lab Med 127, 1603–1605.
- 156. Kew S, Mesa MD, Tricon S, Buckley R, Minihane AM, and Yaqoob P. (2004). Effects of oils rich in eicosapentaenoic and docosahexaenoic acids on immune cell composition and function in healthy humans. Am J Clin Nutr 79, 674–681.
- Kelley DS, Taylor PC, Nelson GJ, and Mackey BE. (1998). Dietary docosahexaenoic acid and immunocompetence in young healthy men. Lipids 33, 559–566.
- 158. Kelley DS, Taylor PC, Nelson GJ, et al. (1999). Docosahexaenoic acid ingestion inhibits natural killer cell activity and production of inflammatory mediators in young healthy men. Lipids 34, 317–324.
- 159. Sydenham E, Dangour AD, and Lim WS. (2012). Omega 3 fatty acid for the prevention of cognitive decline and dementia. Cochrane Database Syst Rev 6:CD005379.
- Bays H. (2007). Fish oil composition of Omacor and the GISSI trial.
 Am J Cardiol 99, 1483–1484.
- Kotani S, Sakaguchi E, Warashina S, et al. (2006). Dietary supplementation of arachidonic and docosahexaenoic acids improves cognitive dysfunction. Neurosci Res 56, 159–164.
- 162. Blyth BJ, Farahvar A, He H, et al. (2011). Elevated serum ubiquitin carboxy-terminal hydrolase L1 is associated with abnormal bloodbrain barrier function after traumatic brain injury. J Neurotrauma 28, 2453–2462.
- 163. Kochanek PM, Berger RP, Bayir H, Wagner AK, Jenkins LW, and Clark RS. (2008). Biomarkers of primary and evolving damage in traumatic and ischemic brain injury: Diagnosis, prognosis, probing mechanisms, and therapeutic decision making. Curr Opin Crit Care 14, 135–141.
- 164. Mehta SS. (2010). Biochemical serum markers in head injury: An emphasis on clinical utility. Clin Neurosurg 57, 134–140.
- 165. Topolovec-Vranic J, Pollmann-Mudryj MA, Ouchterlony D, et al. (2011). The value of serum biomarkers in prediction models of outcome after mild traumatic brain injury. J Trauma 71, S478–486.
- Muller K, Townend W, Biasca N, et al. (2007). S100B serum level predicts computed tomography findings after minor head injury. J Trauma 62, 1452–1456.
- Hodge J, Sanders K, and Sinclair AJ. (1993). Differential utilization of eicosapentaenoic acid and docosahexaenoic acid in human plasma. Lipids 28, 525–531.
- 168. Innis SM. (1992). Plasma and red blood cell fatty acid values as indexes of essential fatty acids in the developing organs of infants fed with milk or formulas. J Pediatr 120, S78–86.
- 169. Spector AA. (2001). Plasma free fatty acid and lipoproteins as sources of polyunsaturated fatty acid for the brain. J Mol Neurosci 16, 159–165; discussion 215–221.
- 170. Carver JD, Benford VJ, Han B, and Cantor AB. (2001). The relationship between age and the fatty acid composition of cerebral cortex and erythrocytes in human subjects. Brain Res Bull 56, 79–85.
- 171. Bouwstra H, Dijck-Brouwer DA, Wildeman JA, et al. (2003). Long-chain polyunsaturated fatty acids have a positive effect on the quality of general movements of healthy term infants. Am J Clin Nutr 78, 313–318.

172. Dunstan JA, Simmer K, Dixon G, and Prescott SL. (2008). Cognitive assessment of children at age 2(1/2) years after maternal fish oil supplementation in pregnancy: A randomised controlled trial. Arch Dis Child Fetal Neonatal Ed 93, F45–50.

- 173. Helland IB, Smith L, Saarem K, Saugstad OD, and Drevon CA. (2003). Maternal supplementation with very-long-chain n-3 fatty acids during pregnancy and lactation augments children's IQ at 4 years of age. Pediatrics 111, e39–44.
- 174. Hibbeln JR, Davis JM, Steer C, et al. (2007). Maternal seafood consumption in pregnancy and neurodevelopmental outcomes in childhood (ALSPAC study): An observational cohort study. Lancet 369, 578–585.
- 175. Innis SM, and Friesen RW. (2008). Essential n-3 fatty acids in pregnant women and early visual acuity maturation in term infants. Am J Clin Nutr 87, 548–557.
- 176. Oken E, Wright RO, Kleinman KP, et al. (2005). Maternal fish consumption, hair mercury, and infant cognition in a U.S. Cohort. Environ Health Perspect 113, 1376–1380.
- Uauy R, and Dangour AD. (2006). Nutrition in brain development and aging: Role of essential fatty acids. Nutr Rev 64, S24

 –33; discussion S72

 –91.
- 178. Williams C, Birch EE, Emmett PM, and Northstone K. (2001). Stereoacuity at age 3.5 y in children born full-term is associated with prenatal and postnatal dietary factors: A report from a population-based cohort study. Am J Clin Nutr 73, 316–322.
- Dullemeijer C, Durga J, Brouwer IA, et al. (2007). n-3 Fatty acid proportions in plasma and cognitive performance in older adults. Am J Clin Nutr 86, 1479–1485.
- 180. Kalmijn S, van Boxtel MP, Ocke M, Verschuren WM, Kromhout D, and Launer LJ. (2004). Dietary intake of fatty acids and fish in relation to cognitive performance at middle age. Neurology 62, 275–280.
- 181. Morris MC, Evans DA, Bienias JL, et al. (2003). Dietary fats and the risk of incident Alzheimer disease. Arch Neurol 60, 194–200.
- 182. Nurk E, Drevon CA, Refsum H, et al. (2007). Cognitive performance among the elderly and dietary fish intake: The Hordaland Health Study. Am J Clin Nutr 86, 1470–1478.

183. Schaefer EJ, Bongard V, Beiser AS, et al. (2006). Plasma phosphatidylcholine docosahexaenoic acid content and risk of dementia and Alzheimer disease: The Framingham Heart Study. Arch Neurol 63, 1545–1550.

- 184. van Gelder BM, Tijhuis M, Kalmijn S, and Kromhout D. (2007). Fish consumption, n-3 fatty acids, and subsequent 5-y cognitive decline in elderly men: The Zutphen Elderly Study. Am J Clin Nutr 85, 1142–1147.
- 185. Bazan NG. (1970). Effects of ischemia and electroconvulsive shock on free fatty acid pool in the brain. Biochim Biophys Acta 218, 1–10
- Neuringer M, Anderson GJ, and Connor WE. (1988). The essentiality of n-3 fatty acids for the development and function of the retina and brain. Annu Rev Nutr 8, 517–541.
- 187. Kim HY, Akbar M, Lau A, and Edsall L. (2000). Inhibition of neuronal apoptosis by docosahexaenoic acid (22:6n-3). Role of phosphatidylserine in antiapoptotic effect. J Biol Chem 275, 35215– 35223.
- Lewis M, Ghassemi P, and Hibbeln J. (2013). Therapeutic use of omega-3 fatty acids in severe head trauma. Am J Emerg Med 31, 273.
- Roberts L, Bailes J, Dedhia H, et al. (2008). Surviving a mine explosion. J Am Coll Surg 207, 276–283.
- Tur JA, Bibiloni MM, Sureda A, and Pons A. (2012). Dietary sources of omega 3 fatty acids: Public health risks and benefits. Br J Nutr 107, S23–52.

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